Waters™

응용 자료

Analysis of Legacy and Emerging Perfluorinated Alkyl Substances (PFAS) in Environmental Water Samples Using Solid Phase Extraction (SPE) and LC-MS/MS

Kenneth J. Rosnack, Douglas M. Stevens, Euan Ross, Kari L. Organtini

Waters Corporation

Abstract

Following the guidance of ISO 25101, achieve detection limits with this method on the Xevo TQ-S micro in compliance with the necessary action levels set by the European Framework Directive and the U.S. EPA health advisory. Analysis of environmental water samples can be accomplished for both legacy and emerging PFASs. The method described in this application note is robust and has been applied to the analysis of a various range of environmental water samples including surface, ground, and waste waters.

Benefits

Performing SPE sample preparation of water samples using the ISO 25101 method for PFAS analysis provides:

- · Highly sensitive analysis using the Xevo TQ-S micro
- · Detection limits in the low to sub- ng/L range to meet regulatory requirements
- · A robust and reliable solution for monitoring PFAS compounds in environmental water matrices

Introduction

Perfluoroalkyl substances (PFAS) are a group of persistent and bioaccumulative anthropogenic pollutants that are common to consumer and industrial processes. They are introduced to the environment through a variety of sources ranging from industrial manufacturing of non-stick coatings to their use in firefighting foams. While this group of compounds encompasses thousands of unique compounds, most advisories currently focus on the two most commonly known, PFOS and PFOA. While there currently are no legal requirements for monitoring of PFASs globally, many countries worldwide do recommend they be monitored at some level. In the United States, the U.S. EPA has set an advisory limit of 70 ng/L (ppt) of total PFOS and PFOA; while in Europe, the European Water Framework Directive has singled out PFOS and its derivatives. The Water Framework Directive is an environmental quality standard and advises an annual average value of 0.65 ng/L for inland surface waters.

To reach detection limits low enough to satisfy advisories, either a highly sensitive mass spectrometer is required, or sample preparation that allows for sample enrichment must be employed. The first option was discussed in a previous application note utilizing the ASTM 7979 procedure with the Xevo TQ-XS.³ This application note will detail the second approach using SPE extraction to enrich water samples with analysis performed on Waters Xevo TQ-S micro. Methodology was adapted from ISO 25101 which was written for analysis of PFOS and PFOA in environmental water samples.⁴ Both approaches are valid options and it depends on a laboratory's resources and testing needs as to which method should be considered.

Experimental

The ISO 25101 method was utilized as a guideline for the sample preparation methodology used for this analysis. Currently, ISO 25101 covers the extraction and analysis of only PFOA and PFOS. For this method, an extended list of PFAS compounds were considered and added. Appendix A contains information on all of the PFAS compounds analyzed in this method, together with a subset of emerging compounds being used to replace the legacy PFAS compounds, including GenX. All standards were obtained from Wellington Laboratories (Guelph, Ontario).

A Certified QC Standard (cat no.: 731) from ERA (Golden, CO), for use with ground and surface waters, was utilized as an instrumental QC check throughout the analysis. The standard contained a mix of 12 PFAS

compounds. Certified values and QC Performance Acceptance Limits for each compound in the mix are provided with the standard, making instrumental QC evaluation quick and straightforward.

Due to widespread use of PFAS substances there are many common sources of potential contamination to the analysis. Since required detection limits are in the low- to sub-ng/L, care must be taken during sample collection, preparation, and analysis. Considering there are many common sources of PFAS contamination in the field and laboratory, it is recommended that any laboratory supplies to be used for this analysis be checked for PFAS contamination before use, as is practical. Contamination is also unavoidable from the chromatographic system. Therefore steps should be taken to minimize any system contribution, and as such, the Waters PFC Analysis Kit (p/n: 176001744) for the UPLC system was utilized. The kit is comprised of PFAS-free components (such as PEEK tubing to replace the conventional Teflon coated solvent lines) and an isolator column that helps to delay any residual background interferences from co-eluting with the analytical peak. Installation of the PFC Analysis Kit is straightforward and quick.⁵ In addition, special mobile phase solvents from Honeywell (Muskegon, MI) were used that were bottled in a manner to reduce residual background PFAS levels.

Sample preparation

Standards were prepared as a mix in methanol and calibration standards were appropriately diluted into 1:1 water:methanol to match the final solvent composition of the samples.

Environmental water samples were collected from various sources including surface water, ground water, influent waste water, and effluent waste water. The surface water and ground water samples were collected locally. Waste water samples were provided by Dr. David Reckhow (University of Massachusetts, Amherst). Samples were collected into pre-washed 250 mL HDPE bottles. A blank of each sample was retained for extraction and the remaining samples were spiked with various levels of PFAS compounds and corresponding isotopically labeled standards. The isotope labeled internal standards were utilized to correct for matrix effects as well as any recovery losses from sample preparation.

Sample extraction was performed using ISO 25101 as a guideline with minor method adjustments to accommodate the extended list of PFAS compounds. Oasis WAX 6 cc, 150 mg SPE Cartridges (p/n: 186002493) were used for the sample extraction of 250 mL water samples. The full method for sample preparation is outlined in Figure 1. This method provides a sample enrichment factor of 250×.

Pre-treatment 1. Adjust pH to <3 2. If sample contains particulates – Filter with glass fiber Pre-treatment 1. 4 mL 0.5% ammonia/methanol solution 2. 4 mL methanol 3. 4 mL water Dry cartridge 3. 4 mL 25 mM acetate buffer (pH 4) 4. Dry cartridge Elute 1. 4 mL methanol – send to waste 2. 8 mL 0.5% ammonia/methanol solution - collect 2. Dilute 200 µL sample in 200 µL 2 mM ammonium acetate 2 mM ammonium acetate

Figure 1. Full method details of SPE sample extraction for water samples.

LC conditions

LC system:	ACQUITY UPLC I-Class PLUS fitted with the PFC Analysis Kit
Column:	ACQUITY UPLC BEH C_{18} 2.1 x 100 mm, 1.7 μm
Column temp.:	35 °C
Sample temp.:	10 °C
Injection volume:	10 μL
Mobile phase A:	95:5 Water:methanol + 2 mM ammonium acetate
Mobile phase B:	Methanol + 2 mM ammonium acetate

Gradient

Time (min)	Flow rate (mL/min)	%A	%В
0	0.3	100	0
1	0.3	80	20
6	0.3	55	45
13	0.3	20	80
14	0.4	5	95
17	0.4	5	95
18	0.3	100	0
22	0.3	100	0

MS conditions

Method events:

MS system:	Xevo TQ-S micro
Ionization mode:	ESI -
Capillary voltage:	0.5 kV
Desolvation temp.:	350 °C
Desolvation gas flow:	900 L/hr
Cone gas flow:	100 L/hr
Source temp.:	100 °C

Divert flow to waste from 16 to 21 minutes

MRM parameters for each compound were optimized using the QuanOptimize tool in MassLynx Software and are listed in Appendix A.

Results and Discussion

Instrument Performance and Detection Limits

The LC-MS/MS method utilized was fit for purpose for the determination of a range of PFAS compounds of interest. An overlay chromatogram showing the chromatography of all the compounds is shown in Figure 2. Peak shape of the early eluting compounds suffer from slight broadening due to the significant difference in solvent composition between the starting LC gradient and sample.

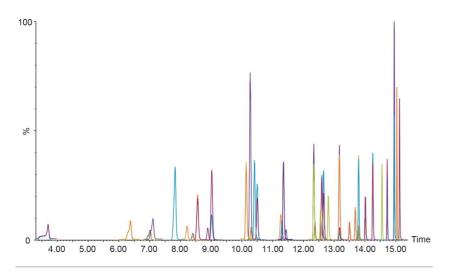


Figure 2. Overlay of all PFAS compounds analyzed in the method.

Detection limits can be seen for all compounds in Table 1. Due to the concentration enhancement provided from the sample preparation procedure, the detection limits are reported as both in-vial and in-sample (250-fold lower than vial concentration) limits. For the most part, in-sample detection limits were sub-ng/L (ppt), reaching to the pg/L (ppq) levels. A few of the less water soluble compounds had ng/L (ppt) detection limits. The detection limits detailed in Table 1 are suitable for current requirements for PFAS testing.

Calibration was very linear over several orders of magnitude for all compounds. An example of a typical solvent calibration curve can be seen in Figure 3, showing an example for PFOA, along with a chromatogram

of PFOA at its detection limit.

During sample analysis, the ERA standard was used as a QC for instrument performance. The instrument performed within the designated Acceptance Limits for all compounds. The average error from the certified values was 15%, although many were below 10% error.

Compound	LOD vial (ng/L)	LOD sample (ng/L)	R²
PFBA	10	0.04	0.999
PFPeA	10	0.04	0.999
PFHxA	10	0.04	0.999
PFHpA	5	0.02	0.999
PFOA	<2	<0.01	0.999
PFNA	10	0.04	0.999
PFDA	10	0.04	0.999
PFUnDA	10	0.04	0.999
PFDoDA	10	0.04	0.999
PFTriDA	10	0.04	0.993
PFTreDA	10	0.04	0.999
PFHxDA	500	2.00	0.994
PFOcDA	2000	8.00	0.988
PFBS	4.4	0.02	0.999
PFPeS	4.7	0.02	0.999
PFHxS	3.7	0.01	0.999
PFHpS	9.5	0.04	0.999
PFOS	3.65	0.01	0.999
PFNS	4.8	0.02	0.999
PFDS	9.6	0.04	0.999
N-EtFOSAA	10	0.04	0.999
N-MeFOSAA	5	0.02	0.999
FHUEA	5	0.02	0.999
FOUEA	5	0.02	0.999
8:2 diPAP	500	2.00	0.997
4:2 FTS	23.4	0.09	0.999
6:2 FTS*	<95	<0.38	0.999
8:2 FTS	9.6	0.04	1.000
PFecHS	9.2	0.04	0.999
FHEA	20	0.08	0.999
FOEA	8	0.03	0.999
FDFA	20	0.08	0.999

- · Achievable detection limits with this method on the Xevo TQ-S micro align with the necessary action levels set by the European Framework Directive and the U.S. EPA health advisory.
- Following the guidance of ISO 25101, analysis of environmental water samples can be accomplished for determination of both legacy and emerging PFASs.
- · The method was verified by the use of the ERA certified QC standard, enhancing confidence in results.
- The method described is robust and has been applied to the analysis of a various range of environmental water samples including surface, ground, and waste waters.

Acknowledgements

The authors would like to acknowledge Honeywell for the productive conversations and considerations for providing the special mobile phase solvents for this work. The authors would also like to acknowledge Dr. David Reckhow and team at the University of Massachusetts, Amherst for providing waste water samples for this work.

References

- 1. EPA PFOA and PFOS Drinking Water Health Advisories. Retrieved 28 November 2018.
- 2. Directive 2013/39/EU of the European Parliament and of the Council of 12 August 2013. Retrieved 28 November 2018.
- K Organtini, G Cleland, and K Rosnack. Large Volume Direct Injection Method for the Analysis of Perfluorinated Alkyl Substances (PFAS) in Environmental Water Samples in Accordance with ASTM 7979-17. Waters Application Note no. 720006329en. June 2018.
- 4. ISO 25101, Water Quality Determination of Perfluorooctansulfonate (PFOS) and Perfluorooctanoate (PFOA) Method for Unfiltered Samples Using Solid Phase Extraction and Liquid Chromatography/Mass Spectrometry. *International Standard*. 2009.
- L Mullin and J Burgess. Ultra Low-Level Detection of Perfluoroalkyl Substances (PFASs) Using the PFC Analysis Kit. Waters Technology Brief 720005701en. May 2016.

Appendix

Compound	CAS number	PFAS class	Type	Precursor	Product	CV	CE	RT	
PFBA	375-22-4	carboxylate	legacy	212.9	169	10	10	3.6	
PFPeA	2706-90-3	carboxylate	legacy	262.9	219	10	5	6.3	
PFHxA	307-24-4	carboxylate	legacy	312.9	269	5	10	8.5	
		22 AUTS			119 319		20 10		
PFHpA	375-85-9	carboxylate	legacy	362.9	169	15	15	10.1	
PFOA	335-67-1	(and and at	lana	412.9	369	10	10	11.0	
PFOA	335-07-1	carboxylate	legacy	412.9	169	10	15	11.3	
PFNA	375-95-1	carboxylate	legacy	462.9	418.9	10	10	12.3	
20.000000	12002 1200				219	838	15	1,575.0	
PFDA	335-76-2	carboxylate	legacy	512.9	468.9 219	15	10	13.1	
		2 2		72223	518.9		10	10272	
PFUnDA	2058-94-8	carboxylate	legacy	562.9	269	25	20	13.8	
PFDoDA	307-55-1	carboxylate	legacy	612.9	568.9	30	10	14.2	
11000/1		out boxy tuto	legacy	0.12.0	169		25		
PFTriDA	72629-94-8	carboxylate	legacy	662.9	618.9	5	10	14.6	
West to West State		The second secon		remedia	169 668.9	- 8	30 15	000000	
PFTreDA	376-06-7	carboxylate	legacy	712.9	169	10	25	14.7	
2511.24	07005.40.5			242.0	768.8		10	45.0	
PFHxDA	67905-19-5	carboxylate	legacy	812.9	169.2	40	40	15.0	
PFODA	16517-11-6	carboxylate	legacy	912.9	868.9	35	15	15.1	
11001	10017-11-0	carboxylate	legacy	312.13	169.2		35	10.11	
PFBS	29420-49-3	sulfonate	legacy	298.9	80.1 99.1	15	30	7.0	
			Section and the second		80.1		30	000000	
PFPeS	2706-91-4	sulfonate	legacy	348.9	99.1	10	30	8.8	
10.000,000,000	2000 000 000 000 000 000		2000000000	(App. 100 p.	80.1	V22744V	35	11300250	
PFHxS	3871-99-6	sulfonate	legacy	398.9	99.1	10	30	10.3	
DEUne	375-92-8	sulfonate	logocy	448.9	80.2	15	35	11.4	
PFHpS	3/5-92-6	suiionate	legacy	446.9	99.1	15	35	11.4	
PFOS	1763-23-1	sulfonate	legacy	498.9	80.2	15	40	12.3	
					99.1 80.2		40 40		
PFNS	N/A	sulfonate	legacy	548.9	99.2	20	40	13.2	
DEDO	225 77 2		taman.	F00.0	80.2	05	40	10.0	
PFDS	335-77-3	sulfonate	legacy	598.9	99.1	25	40	13.8	
N-MeFOSAA	2991-50-6	sulfonamidoacetic acid	legacy	569.9	418.9	35	20	13.5	
				879,5.50.2°	219.1	10000	25	Charles	
N-EtFOSAA	2355-31-9	sulfonamidoacetic acid	legacy	584	418.8 525.9	15	20	13.8	
			•		292.9		10		
FHUEA	70887-88-6	unsaturated telomer acid	legacy	356.9	243	10	35	10.4	
FOUEA	70887-84-2	unsaturated telomer acid	legacy	456.9	393	10	10	12.6	
TOOLA	70007-04-2	unsaturated telemer dela	legacy	450.5	343	10	40	12.0	
8_2 diPAP	678-41-1	phosphate ester	legacy	989	97 542.5	10	20	15.0	
V (1000)			140000000000000000000000000000000000000		307	70.50.00	15		
4_2 FTS	757124-72-4	telomer sulfonate	legacy	326.9	81.1	15	35	8.4	
					407		20		
6_2 FTS	29420-49-3	telomer sulfonate	legacy	426.9	344.9	15	10	11.3	
					81		35		
8_2 FTS	39108-34-4	telomer sulfonate	legacy	526.9	506.8 444.6	15	25 10	13.1	
0_2110	00100 01 1	tolollol ballollato	legacy ozolo	.cgacy	02010	81.2		40	1011
PFecHS	67584-42-3	cyclic	logogy	460.9	380.9	40	30	11.2	
Precns	0/304-42-3	Cyclic	legacy	400.9	99.1	40	30	11.2	
FHEA	53826-12-3	telomer acid	legacy	376.9	292.9	5	15	10.5	
				1000000	313 393	8	5 10	100000	
FOEA	27854-31-5	telomer acid	legacy	476.9	413	5	5	12.6	
EDE 1	E2020 12 1	halaman and d	laa	F70.0	492.9		15		
FDEA	53826-13-4	telomer acid	legacy	576.9	512.9	15	5	14.0	
FHpPA	812-70-4	other	legacy	440.9	336.9	15	10	12.5	
			57		317		20	/ (TO 100)	
ADONA	958445-44-8	other	emerging	376.9	251 85	10	10 25	10.2	
//					350.9		25		
9CI-PF3ONS	73606-19-6	other	emerging	530.9	83	15	25	12.8	

Appendix 1

Compound	CAS number	PFAS class	Туре	Precursor	Product	CV	CE	RT
11CI-PF3OUdS	73606-19-6	other	emerging	630.9	450.8	30	30	14.0
					83 119		30 35	
GenX	13252-13-6	other	emerging	285	185	- 5	7	9.0
PFMBA	863090-89-5	other	emerging	278.9	85	10	10	7.0
	000000 00 0	ounci	omorging	27010	235		5	,,,
NFDHA	151772-58-6	other	emerging	294.9	85 201	- 5	10	8.2
PFEESA	113507-82-7	other		314.9	83	15	20	7.8
		VI 10 (20 m) (20)	emerging	***************************************	135	15	20	0.000
13C-PFBA 13C5-PFPeA	-	-		216.9 267.9	172 223	10	10 5	3.6 6.3
					272.9		5	
13C5-PFHxA	1.5	-	S=	317.9	119.9	10	20	8.5
					321.9		10	22.5
13C4-PFHpA	-	-	_	366.9	169 172	15	15 15	10.1
					375.9	. 0=1	10	
13C8-PFOA	10 .70		(25)	420.9	172	- 5	15	11.3
13C9-PFNA	-	_	-	471.9	426.9	10	10	12.3
					223 473.9	0.004	15 10	20102000
13C6-PFDA	-	-		518.9	223	5	15	13.1
13C7-PFUnDA	_	_	_	569.9	524.9	- 5	10	13.8
1007 TT OHDA	= =		=======================================	500.0	274		15	10.0
13C-PFDoDA	12	_	-	614.9	569.9 169	10	10 25	14.2
100 11 202/1				01410	269.1		20	13114
13C2-PFTreDA		_	_	714.9	669.9	25	10	14.7
1002 11 11007		*****		7,14.5	169		35	1711
13C2-PFHxDA	-	_	-	815	769.9 169.3	30	15 35	15.0
12C2 DEBC	1000	KIK	657	201.0	80	10	30	7.0
13C3-PFBS	-	-	** <u>*</u>	301.9	99	10	25	7.0
13C3-PFHxS	-		=	401.9	80.1 99.1	10	40 35	10.3
					80.1		40	720 minutes
13C8-PFOS	-	-	1000 T	506.9	99.1	15	40	12.3
D5-N-EtFOSAA	-	-	-	589	418.9	- 30	20	13.8
					506.9 418.9		15 20	
D3-N-MeFOSAA	_	_	_	572.9	482.7	35	15	13.5
				****************	514.7		20	10.000
13C-FOUEA	1. 	-	-	458.9	393.9	25	10	12.6
					119.1 97.3		40	
13C4-8:2 diPAP	-	=	e =	993	544.8	30	25	15.0
13C2-4:2 FTS	-	-	_	328.9	308.9	40	15	8.4
					81		25	
13C2-6:2 FTS	-	-	-	428.9	367 408.8	10	20	11.3
13C2-8:2 FTS	12		-	528.9	508.9	10	20	13.1
1302-0.2 F13				320.9	81	10	35	13.1
13C-FHEA	-	5	-	378.9	293.9 64.1	- 5	10 	10.5
100 5051				170.0	393.9		15	40.0
13C-FOEA	6.50		8 .7 v	478.9	64.1	10	10	12.6
13C-FDEA	-	_	-	578.9	493.9	- 25	5	14.0
AND				No. of the last of	64.2 169		5 12	2001A)
13C3-GenX	-	-	-	287	119	- 5	12	9.0
					NAME OF TAXABLE PARTY.			

Featured Products

ACQUITY UPLC I-Class PLUS System https://www.waters.com/134613317

Xevo TQ-S micro Triple Quadrupole Mass Spectrometry https://www.waters.com/134798856

MassLynx https://www.waters.com/513662

Available for purchase online

ACQUITY UPLC BEH C18 Column, 130Å, 1.7 μ m, 2.1 mm X 100 mm <

https://www.waters.com/waters/partDetail.htm?partNumber=186002352>

Oasis WAX 6 cc Vac Cartridge, 150 mg Sorbent per Cartridge, 30 µm Particle Size <

https://www.waters.com/waters/partDetail.htm?partNumber=186002493>

PFC Analysis Kit https://www.waters.com/waters/partDetail.htm?partNumber=176001744>

720006471, January 2019

©2019 Waters Corporation. All Rights Reserved.