

アプリケーションノート

Analysis of Plant Alkaloids Through Accurate Mass Screening and Discovery

Jeff Goshawk, Michelle Wood

日本ウォーターズ株式会社



For forensic toxicology use only.

Abstract

In this study, the Forensic Toxicology Application Screening Solution with UNIFI was applied to a selection of plant alkaloids. The ease by which the scientific library items can be created and updated has been clearly demonstrated. The UNIFI Scientific Information System v1.8 was used to process the MS^E data and for these plant alkaloids multiple adducts were detected. The fragment match functionality was also able to assign sub-structures to high-energy ions. Additionally, the new discovery tool has been shown to enhance the elucidation of unknown components.

Benefits

Analyze plant alkaloids using the Forensic Toxicology Application Screening Solution with UNIFI¹ to demonstrate the simplicity of library creation and expansion. This application note also showcases the power of the latest suite of discovery tools within the UNIFI Scientific Information System v1.8.

Introduction

Over the last decade there has been a significant increase in the popularity of time-of-flight mass spectrometry (Tof-MS) for multi-residue analysis. Accurate mass imparts high specificity for substance identification and, together with the isotopic data, can provide the user with the opportunity to propose likely elemental compositions. The proposal of elemental formulae is often the starting point for a complex multi-stage process to elucidate chemical structures.

For screening, accurate mass instrumentation presents a significant, and key, advantage over its nominal mass counterpart, i.e., an ability to implement screening methodologies without the requirement of reference material. In this particular workflow the theoretical (expected) exact mass can be determined empirically from the elemental formula. In a toxicological setting this can provide a valuable means with which the analyst may 'prospectively' target novel psychoactive drugs, or new substances and metabolites where reference material may not yet, be available.

An on-going initiative to expand the UNIFI Toxicology Scientific Library led to the analysis of a series of plant alkaloids. These nitrogen-containing compounds are derived from plants and plant material. They are pharmacologically active and have been used for many centuries for both medicinal and recreational purposes. Consequently, their analysis is of forensic importance. Analysis of these substances provided an opportunity to evaluate the tools within the UNIFI Scientific Information System for both target assignment and structural elucidation.

ACQUITY UPLC conditions

System:	ACQUITY UPLC I-Class (FTN)
Column:	ACQUITY HSS C_{18}, 2.1 x 150 mm, 1.8 μm
Run time:	15 min
Vials:	Waters Maximum Recovery Vials
Column temp.:	50 °C
Sample temp.:	10 °C
Injection vol.:	10 µL
Flow rate:	0.4 mL/min
Mobile phase A:	5 mM aqueous ammonium formate, adjusted to pH 3.0
Mobile phase B:	Acetonitrile containing 0.1% formic acid

Gradient:

Time	%A	%В
0.00	87	13
0.50	87	13
10.00	50	50
10.75	5	95
12.25	5	95
12.50	87	13
15.00	87	13

MS conditions

MS system:	Xevo G2-S QTof
Ionization mode:	ESI+
Source temp.:	150 °C
Desolvation temp.:	400 °C
Desolvation gas:	800 L/h
Reference mass:	Leucine enkephalin [M+H] ⁺ = <i>m/z</i> 556.2766
Acquisition range:	m/z 50–1000
Scan time:	0.1 s
Capillary voltage:	0.8 kV
Cone voltage:	25 V
Collision energy:	Function 1: 6 eV
	Ramped 10 to 40 eV
Data management	Forensic Toxicology Screening Application Solution with UNIFI v1.8

Experimental

Materials

The following plant alkaloids were obtained from Sigma-Aldrich (Poole, UK) as solid material: amygdalin, berberine chloride, bufalin, coumarin, digitoxin, gitoxin, lanatocide C, neriifolin, and α -solanine.

Sample preparation

Individual stock solutions of the plant alkaloids were initially prepared, by dilution with methanol, to a concentration of 10 μ g/mL; these solutions were stored at -20 °C until further use. Prior to Tof-MS analysis, the stock solutions were further diluted with mobile phase A to yield samples for injection at a concentration of 1 μ g/mL.

Results and Discussion

Prior to analysis, a new UNIFI Scientific Library was created specifically for plant alkaloids, by simply entering the names of the nine alkaloids. A MOL file describing the structure of each substance was added to each entry in the library (Figure 1). Individual solutions of the plant alkaloids were injected and data were acquired using the standard screening conditions supplied with the Forensic Toxicology Screening Application Solution with UNIFI.¹ These data were subsequently processed using the UNIFI Scientific Information System and screened against the new plant alkaloid library.

tem description UPAC name formula C30H4608 full formula C30H4608 full south rease 534.6814 donoiutopic mass 534.3193 tem tag	roperty	Value	Specify item name and library
VPAC name formula C30H4608 fill formula C30	tem type	Compound	0-40
UPAK name ommula C30H4608 liffomula C30H4608 vonge molar mass S34.8814 nonoixotopic mass S34.3193 em tag	tem description		
C3014603 C3014603 4III formula C304603 Average molar mass 534.6814 Monisotopic mass 534.383 tem tag Image: California formula	UPAC name		\checkmark
fill formula C3014608 Wersge molar mass 534.6814 Wonoiotopic mass 534.3193 tent tag Image: California State of Californi State of California State of Cal	Formula	C30H46O8	
Werage molar mass 534.6814 Wonoisotopic mass 534.3193 tem tag	Hill formula	C30H46O8	
tem tag	Average molar mass	534.6814	
	Monoisotopic mass	534.3193	Remove explicit hydrogens
	Item tag		0 Viewer properties

Figure 1. Creating a library entry for neriifolin. Existing MOL file structures can be appended (Load structure) or created by standard chemical drawing packages and subsequently appended (New structure).

Identification through the application of in-silico fragmentation techniques

The presence of each plant alkaloid was confirmed through the mass accuracy of the protonated precursor ion in combination with theoretical fragment ions that were automatically generated from the structure of each substance and matched to ions in the high-energy spectrum.

Figure 2 shows the identification of α -solanine as presented in UNIFI. The Component Summary table presents the information related to the identification of this alkaloid and includes; the observed m/z value together with the deviation from the expected m/z value, the difference between measured and theoretical isotope patterns in terms of both m/z and intensity distributions, the observed retention time, the number

of theoretical fragment ions found, and the detector counts, which represents the abundance of all the lowenergy ions associated with the detected compound.

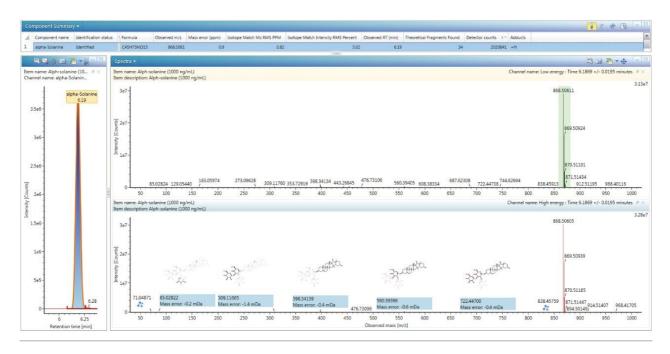


Figure 2. Identification of a-solanine in the UNIFI Scientific Information System.

Updating library entries

All of the alkaloids were identified on the basis of the mass accuracy of the precursor ion and theoretical fragment ions generated during processing. Upon identification, a retention time was associated with each substance. With UNIFI, the library entries can be updated directly from the analysis such that they contain the expected retention time and the expected *m/z* value for each assigned adduct and fragment ion. Following the update, a typical library entry has information similar to that shown for neriifolin in Figure 3. This additional information can be used to target the substance in subsequent analyses.

Nenifo	olin (Plant Alk	kaloids]			 						🏠 Tools 🔻
operty			Value							0	
m type			Compound								
m descr	ription									0	
IPAC nar	me										
ormula			C30H46O8							- Y	
ill formu	ıla		C30H46O8							A Marine H	
verage n	molar mass		534.6814					Ŧ			
Ionoisote	opic mass		534.3193					0. I			
								0 mm	0	H	
	on results 🕶							K.			l
dd Ed	dit Delete Priority 1 + In		Formula	Neutral Mass (Da)			Expected m/z	Expected RT (min)	Ionization technique	Detail type Mise	
dd Ed	dit Delete Priority 1 + In 1	5769486		Neutral Mass (Da) 534.319	1	None	Expected m/z 535.3265	Expected RT (min) 9.300	ESI+	MSe	
dd Ea	dit Delete Priority 1 + In 1 5	5769486 1166637	C23H31O2		1	None	Expected m/z 535.3265 339.2319	Expected RT (min) 9.300 9.300	ESI+	MSe MSe	E
dd Eo	dit Delete Priority 1 + In 1	5769486 1166637 864505			1	None	Expected m/z 535.3265	Expected RT (min) 9.300 9.300 9.300	ESI+	MSe	E
dd Eo	dit Delete Priority 1 - In 1 5 6	5769486 1166637 864505 676380	C23H31O2 C23H35O4		1 1 1	None CID CID	Expected m/z 535.3265 339.2319 375.2530	Expected RT (min) 9.300 9.300 9.300 9.300	ESI+ ESI+ ESI+	MSe MSe MSe	Ę
dd Eo	dit Delete Priority 1 - In 5 6 7	5769486 1166637 864505 676380 308892	C23H31O2 C23H35O4 C23H33O3		1 1 1 1	None CID CID CID	Expected m/z 535.3265 339.2319 375.2530 357.2424	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300	 ESI+ ESI+ ESI+ ESI+ 	MSe MSe MSe	[
dd Eo	dit Delete Priority 1 + In 5 6 7 8	5769486 1166637 864505 676380 308892	C23H31O2 C23H35O4 C23H33O3 C4H5O2 C30H45O7		1 1 1 1	None CID CID CID CID CID	Expected m/z 535.3265 339.2319 375.2530 357.2424 85.0284	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300	 ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ 	MSe MSe MSe MSe	
dd Eo	dit Delete Priority 1 1 In 5 6 7 8 9	5769486 1166637 864505 676380 308892 131214 117844	C23H31O2 C23H35O4 C23H33O3 C4H5O2 C30H45O7		1 1 1 1 1 1	None CID CID CID CID CID CID CID	Expected m/z 535.3265 339.2319 375.2530 357.2424 85.0284 517.3160	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	 ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ 	MSe MSe MSe MSe MSe	
dd Ea	dit Delete Priority 1 In 5 6 7 8 9 10	5769486 1166637 864505 676380 308892 131214 117844 108359	C23H31O2 C23H35O4 C23H33O3 C4H5O2 C30H45O7 C5H7O2		1 1 1 1 1 1 1	None CID CID CID CID CID CID CID	Expected m/z 535.3265 339.2319 375.2530 357.2424 85.0284 517.3160 99.0441	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+ ESI+	MSe MSe MSe MSe MSe MSe MSe	
dd Eo	dit Delete Priority 1 In 5 6 7 8 9 10 11	5769486 1166637 864505 676380 308892 131214 117844 108359 91115	C23H31O2 C23H35O4 C23H33O3 C4H5O2 C30H45O7 C5H7O2 C30H43O6		1 1 1 1 1 1 1 1	None CD CD CD CD CD CD CD CD CD	Expected m/z 538.3265 339.2319 375.2530 357.2424 85.0284 517.3160 99.0441 499.3054	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	ESI+	MSe MSe MSe MSe MSe MSe MSe MSe	
dd Eo	dit Delete Priority 1 1 5 6 7 8 9 10 11 11 12	5769486 1166637 864505 676380 308892 131214 117844 108359 91115 82439	C23H31O2 C23H35O4 C23H35O3 C4H5O2 C30H45O7 C5H7O2 C30H43O6 C15H19O2		1 1 1 1 1 1 1 1 1	None CD CD CD CD CD CD CD CD CD CD CD CD CD	Expected m/z 535.3265 338.2319 375.2530 357.2424 85.0284 517.3160 99.0441 499.3054 231.1380	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+ 651+	MSe MSe MSe MSe MSe MSe MSe MSe MSe	
dd Eo	dit Delete Priority 1 + In 5 6 7 8 9 10 11 11 12 13	5769486 1166637 864505 676380 308892 131214 117844 108359 91115 82439 52590	C23H31O2 C23H35O4 C23H33O3 C4H5O2 C30H45O7 C5H7O2 C30H43O6 C15H19O2 C6H9O3		1 1 1 1 1 1 1 1 1 1	None CD CD CD CD CD CD CD CD CD CD CD CD CD	Expected m/z 535.3265 339.2319 375.2530 357.2424 85.0284 517.3160 99.0441 499.3054 231.1380 129.0546	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	+23 +23 +23 +23 +23 +23 +23 +23 +23 +23	MSe MSe MSe MSe MSe MSe MSe MSe MSe MSe	
dd Ed	dit Delete 1 ∩ 1 3 5 7 8 9 10 11 12 13 14	5769486 1166637 864505 676380 308892 131214 117844 108359 91115 82439 52590	C23H3102 C23H3504 C23H3303 C4H502 C30H4507 C5H702 C30H4306 C15H1902 C6H903 C6H702 C4H702		1 1 1 1 1 1 1 1 1 1 1 1	None CD CD CD CD CD CD CD CD CD CD CD CD CD	Expected m/z 535.3265 339.2319 375.2530 357.2424 85.0284 517.3160 919.0441 499.3054 223.1380 129.0546 131.0441	Expected RT (min) 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300 9.300	ESI- ESI- ESI- ESI- ESI- ESI- ESI- ESI-	MSe MSe MSe MSe MSe MSe MSe MSe MSe MSe	

Figure 3. Library entry for neriifolin. The lower section of the composite is now populated with expected retention time and the expected m/z values of precursor and fragment ions.

Multiple adducts

Data for gitoxin, one of the other alkaloids investigated in this study, is shown in Figure 4. The low-energy ions assigned to this substance are highlighted in green within the spectrum and correspond to the protonated isotope cluster. The detector counts determined for the protonated isotope cluster of gitoxin is 568. The high-energy spectrum is annotated with sub-structures of gitoxin, as determined automatically by UNIFI and associated to the high-energy spectral peaks as fragment ions.

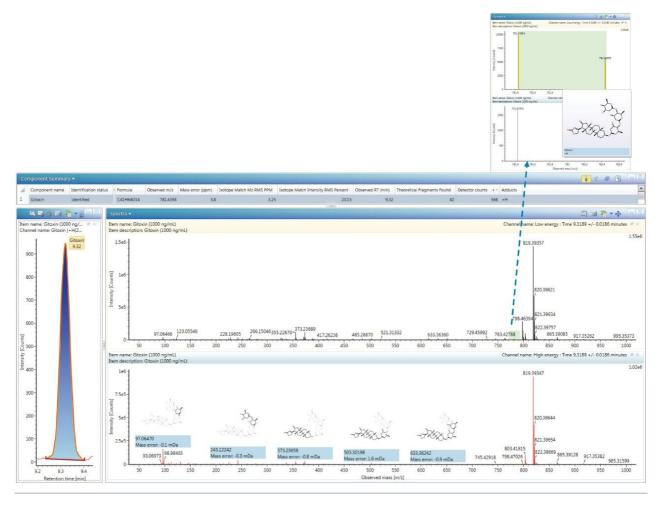


Figure 4. Identification of gitoxin in the UNIFI Scientific Information System.

Further examination of the low-energy spectrum for this substance revealed that some of the ions may correspond to other adducts of gitoxin. Consequently the data was reprocessed to target the $[NH_4]^+$, $[Na]^+$, and $[K]^+$ adducts in addition to the protonated species. Figure 5 details the isotope clusters in the low-energy data assigned to each adduct following reprocessing. The assignment of the additional adducts to gitoxin has been reflected in the detector counts which has increased from 568, determined from the isotope cluster of the protonated adduct, to 118680. Similar results were obtained for the other substances in this analysis.

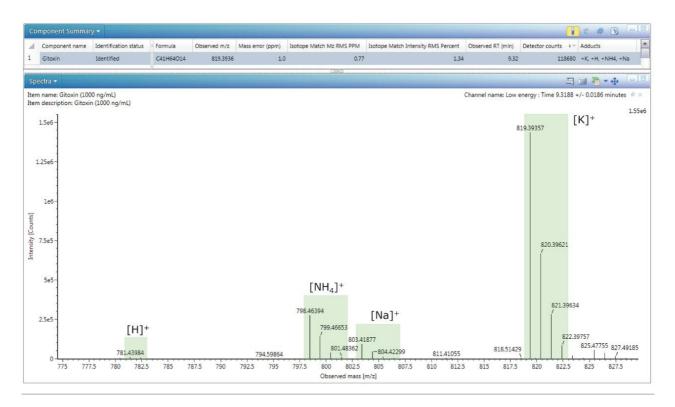


Figure 5. Multiple adduct assignment for gitoxin

The discovery tool

Another new feature in the UNIFI Scientific Information System v1.8 is the discovery tool, which chains together elemental composition, library searching and fragment match functionality into a single step process making it easier to obtain the identity of unexpected substances within a sample. The parameters used to run the discovery tool are detailed in Figure 6A–D.

The first set of parameters, displayed in Figure 6A, control the maximum number of elemental compositions returned for each component, and the number of library hits returned for each elemental composition. For each component selected, the measured m/z value is submitted to the elemental composition application, the parameters of which are displayed in Figure 6B. Each scientific formula returned by the elemental composition application is then automatically submitted to the list of selected libraries. The libraries can either belong to the UNIFI Scientific Library or, if connected to the internet, ChemSpider. The dialog showing the selection of ChemSpider libraries is presented in Figure 6C.

Every hit for each scientific formula that is returned from the library search is then automatically submitted to the fragment match application, provided the library hit has an associated structure in the form of a MOL file.

The fragment match application performs a systematic bond disconnection of each structure, applying the

parameters selected through the dialog displayed in Figure 6D, and matches the *m/z* values of theoretical sub-structures to measured high-energy fragment ions. The number of fragment ions matched and the percentage of the intensity of the high-energy spectrum accounted for by those matches are both determined.

А

Discovery					(0
Paramete	rs				4
Discovery	Elemental Composition	ChemSpider	Fragment Match		
Elemental	Composition		ChemSpider	Scientific Library	
Minimum	Minimum i-FIT Confidence: 10 %		Minimum citations:	0	
Number o	of compositions:	5	Number of hits:	50	
) Start	Cancel				
3					
Discoverv	-				

Discovery Eleme	ental Composition	ChemSpider	Fragment M	atch			
Composition				m/z Tolerance:	2	mDa	🔽 Use Senior rule
	ments selection			Electron state:	Even	•	Use Carbon/Hydrogen ratio filter
Select elements Use formula from parent							🔽 Use Carbon/Hetero-atom ratio file
Selected elemen	ts: C, H, N, O, S, 0	CI, Br		Minimum DBE:	-1	🔽 Use multi-atom filter	
Adducts				Maximum DBE:		50	
Automatic ad Selected adduct:		Calact	idduct	Number of isotopes before selected peak:		0	
Total adducts ch	413	1	100903	Number of isotopes to use:		3	

Parameters	
Discovery Elemental Composition ChemSpider Fragment Match	
Available libraries: Selecte	ed libraries:
A&J Pharmtech FDA L	JNII - NLM
A1 BioChem Labs	
A2Z Chemical	
Abacipharm	
Abblis Chemicals	
Abcam	
ABI Chemicals	
Abmole Bioscience	

D

Parameter	rs									1
Discovery	Elemental Co	mposition	ChemSpider	Fragment Match						
🗸 Use sm	artsScores	Multiple:	4	Alpha:	5	DBE minimum:	-1.5	Mode: Auto	omatic •	
Phenyl:	8	Other:	1	Hydrogen difference:	6	DBE maximum:	50	Filter peaks by	intensity	
Aromatic:	6	Bonds:	4	Allow scores below:	8	Neutral:	On •	Number of peaks:		
Ring:	2	Hetero:	0.5	Delta (mDa):	2	H Penalty:	0			

Figure 6. Discovery tool in UNIFI. A) General discovery tool parameters. B) Elemental composition parameters. C) ChemSpider parameters. D) Fragment match parameters.

For the purposes of illustration, the candidate component identified as amygdalin in the targeted analysis was submitted to the discovery tool. The results, upon running the application with respect to the parameters shown in Figure 6A–D, are presented in Figure 7.

The component submitted to the discovery tool was Candidate Mass m/z 458.1649. The results show that one elemental composition, $C_{20}H_{27}NO_{11}$, with an i-FIT confidence of 89% was determined for m/z 458.1649. This elemental composition, was automatically submitted to the FDA UNII – NLM library within ChemSpider and a hit for amygdalin was returned with a list of synonyms, a structure and the number of citations. The structure was used automatically in conjunction with fragment match and appropriate sub-structures were assigned to the high-energy spectrum associated with Candidate Mass m/z 458.1649, as shown in Figure 7. The number of high energy fragments matched by sub-structures and the percentage of the intensity of the high energy spectrum accounted for by those fragment matches are displayed for the library hit.

Access to this information for a range of components, elemental compositions, and library hits enables the analyst to make an informed decision with respect to the identity of unexpected substances in their samples.

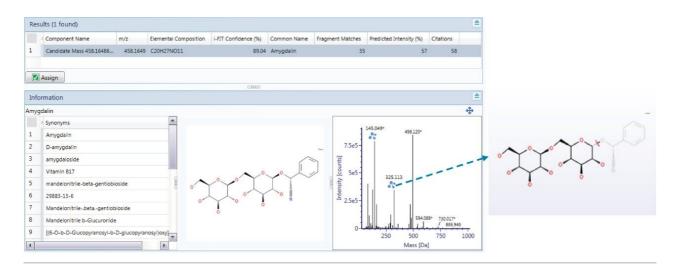


Figure 7. A typical result in the discovery tool.

Conclusion

In this study, the Forensic Toxicology Application Screening Solution with UNIFI¹ was applied to a selection of plant alkaloids. The ease by which the scientific library items can be created and updated has been clearly demonstrated. The UNIFI Scientific Information System v1.8 was used to process the MS^E data and for these plant alkaloids multiple adducts were detected. The fragment match functionality was also able to assign sub-structures to high-energy ions. Additionally, the new discovery tool has been shown to enhance the elucidation of unknown components.

References

1. Forensic Toxicology Screening Application Solution. Waters Brochure (P/N 720004830EN).

Featured Products

ACQUITY UPLC I-Class PLUS System https://www.waters.com/134613317 Forensic Toxicology Screening Application Solution with UNIFI https://www.waters.com/134779723

Available for purchase online

ACQUITY UPLC HSS C18 Column, 100Å, 1.8 μm, 2.1 mm X 150 mm, 1/pkg < https://www.waters.com/waters/partDetail.htm?partNumber=186003534>

720005461, July 2015

©2019 Waters Corporation. All Rights Reserved.